

The Determination of Normal Percentages of Syncytiotrophoblastic Knots in Various Regions of Placenta: Where to Count the Syncytial Knots

Sinsityotrofoblastik Düğümlerin Plasentanın Farklı Bölgelerindeki Normal Yüzdelerinin Belirlenmesi: Sinsityal Düğümleri Nereden Sayalım

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ABSTRACT

Objective: The marginal, basal and subchorial regions of the placenta are considered to be more hypoxic than other regions. Therefore, it is not recommended to determine the increase in syncytiotrophoblast knots, based on the major morphological change in placental hypoxia, from the samples taken from these regions. However, the normal count of knots at various regions of placenta is not investigated.

Material and Method: In this study we have sampled morphologically and clinically normal placenta with eccentric cord insertion from various sites, either close to cord entrance or away from it (marginal, non-marginal basal, non-marginal subchorial, and nonmarginal midparanchymal). The number of knots was calculated on a total of at least 100 villi for each placental sample. The normal amount of knots in different regions and comparison between them were investigated. Twenty-eight placentas with eccentric cord insertion were sampled in the same manner. Hot spots from the above mentioned regions were counted in a total of 100 villi.

Results: No significant difference was found between the dual comparison of the mean percentages of different regions (p: 0.148). The variety of hypoxia in different regions of the placenta could not be demonstrated in this study.

Conclusion: It is found that there is no difference in perfusion that can be morphologically demonstrated with increase in syncytiotrophoblast knot, between different regions of placenta.

Key Words: Hypoxia, Placenta, Syncytiotrophoblast

ÖZ

Amaç: Plasentanın marjinal, bazal ve subkoryal bölgelerinin diğer bölgelerine göre daha hipoksik olduğu düşünülür. Buna göre, plasental hipoksideki morfolojik bulgular temelinde sinsityotrofoblastik düğümlerdeki artışın bu bölgelerden alınacak örneklerden belirlenmesi önerilmez. Her şeye karşın plasentanın farklı bölgelerindeki düğümlerin normal sayısı incelenmemiştir.

Gereç ve Yöntem: Bu çalışmada, eksantrik kord girişli, morfolojik ve klinik açıdan normal plasentaların kord girişine yakın veya uzak, farklı alanlarından (marjinal, non-marjinal bazal, non-marjinal-subkoryal, non-marjinal midparenkimal) örnekler alındı. Düğümler, her bir plasental örnekte total en az 100 villüsten sayıldı. Farklı bölgelerdeki normal düğüm sayısı incelendi ve karşılaştırma yapıldı. Eksantrik kord girişli 28 plasenta benzer biçimde örneklendi. Yukarıda belirtilen bölgelerdeki sıcak noktalar total 100 villüsten sayıldı.

Bulgular: Farklı bölgelerin ortalama yüzdeleri dual karşılaştırıldığında anlamlı bir fark saptanmadı (p: 0,148). Bu çalışmada, plasentanın farklı bölgelerindeki hipoksi değişiklikleri gösterilemedi.

Sonuç: Yazarların düşüncesine göre plasentanın farklı bölgelerinde sinsityotrofoblastik düğüm artışı ile morfolojik olarak gösterilebilecek perfüzyon farklılıkları bulunmamaktadır.

Anahtar Sözcükler: Hipoksi, Plasenta, Sinsityotrofoblast

INTRODUCTION

Tenney Parker Changes (TPC) are the major marker of maternovascular perfusion insufficiency. They are characterized by syncytial knot increase and villus clustering. Syncytial knot is a marked clustering of the svncvtial nuclei under the light microscope. Studies performed with the electron microscope demonstrate various types of syncytial knots with different mechanisms of formation. These formation types usually cannot be differentiated under the light microscope. The syncytial knot increase has histomorphological significance (1). There is ongoing research among pathologists to designate the most appropriate definition of syncytial knots (2). The existence of syncytial knots in more than one third of the villi is defined as TPC. It is recommended to calculate the knot number from the midparenchymal region, because the basal, marginal and the subchorial regions are considered to be more hypoxic than the other regions (3). However, there is no standard regarding the normal number of knots from different regions. This study attempts to determine the normal amount of syncytial knots in different regions and to demonstrate whether a difference exists between them.

MATERIALS and METHODS

Twenty-eight placentas received at Ağrı Women and Children's Hospital, Department of Pathology were chosen for the study. All the samples were term gestational placentas with eccentric cord entrances. The weight of the placentas varied between 414 and 470 grams, which was within normal limits (4). They were all reported as late 3rd trimester placentas that were morphologically normal. Descriptive additional diagnoses coexisted which did not affect the final diagnosis. The following cases thought to bias the study either clinically or morphologically were not included in the study; • Cases with no eccentric entrances: central, marginal and other cord entrances were thought to have different perfusions than eccentric ones.

• Placenta-induced hypertension (PIH): It is reported that there is no uniform physiologic change in different vessels of decidua basalis and in various regions of a single vessel as well (3). That is why it is accepted that even normal placentas may demonstrate physiological conversion insufficiency that is frequently seen in cases with preeclampsia and SGA (Small for Gestational Age) (5). As the deficiency of physiological conversion may increase the perfusion variances in the placenta, PIH cases with normal morphology were excluded from the study.

• Cases with villous maturation score of other than 22 and 23: Villous maturation scores are included in the reports of our department. The scores of 22 and 23 are defined as normal late third trimester placentas as demonstrated by Benirske and Kaufmann (1) and shown in Figure 1 and Figure 2.

• Cases with chorioamnionitis: The endothelin amount is increased in chorioamnionitis which in turn results in fetal vasoconstriction and hypoperfusion (6). VEGF is a growth factor increasing inflammatory cell migration. Its expression is increased both in hypoxic cases and in chorioamnionitis (7,8). Chorioamnionitis cases were therefore excluded from the study even with scores of 22 and 23.

• Cases with morphological changes associated with hypoxia and decreased perfusion: TPC is not the single finding of hypoxia and cases with the following changes demonstrated even focally were therefore excluded from the study: Increased intervillous fibrin, distal villous hypoplasia, acute atherosis, mural hypertrophy of membrane arterioles, muscularised basal plate arteries, increased placental side giant cells, increased immature intermediate trophoblasts,



Figure 1: Score 22 (Villous maturation score) (H&E; x100).



Figure 2: Score 23 (Villous maturation score) (H&E; x100).

thin umbilical cord, laminar necrosis, and microscopic chorionic pseudo cysts (2, 9, 10).

Placentas were kept in a solution of 10% formaldehyde for a night and were cut into two segments from the cord entrance parallel to its longitudinal axis (Figure 3). Six blocks were taken from each case. The sampling regions were as follows;

- 1X: Marginal zone close to cord insertion.
- 1: Non-marginal zone close to cord insertion.
- 2: Non-marginal zone far from cord insertion.
- 2X: Marginal zone far from cord insertion (Figure 4).
- O: Fetal membranes and cord.
- T: Horizontally sampled placental base.

After this sampling, the placentas were dissected parallel to the first incision and additional samples were taken from



Figure 3: The dissection of placenta in this study.



Figure 4: Illustration of zones sampled in the study. 1X, marginal zone close to cord insertion; 2x, marginal zone far from cord insertion; 1, central zone close to cord insertion; 2, central zone far from cord insertion.

regions with an unusual appearance. In order to increase the standardization in perfusion, the central regions of maternal cotyledons at the opening site of the uterine arteries were chosen. All the samples were cut in 5 micron thickness and stained with hematoxylin and eosin. The pathologist investigated the samples. All the counting was done from the hot spots and knot numbers were determined from at least 100 villi for each dissected sample. Each syncytial knot and syncytial knot bridging was calculated as 1 point. During these scorings, accumulations with crowded nuclei, which are easily seen in 10x magnifications, were counted without question. For smaller knots, the inclusion presence of at least 6 nuclei was required. The compact accumulation of the nuclei was stipulated. Only the knots on terminal villi were counted. Syncytial knots existing on more than 1 villus due to slicing were also included in the study if they met the criteria above. Each syncytial knot was scored as 1. Syncytial knots including at least 6 nuclei and combining more than two villi were defined as syncytial knot bridging. Each syncytial knot bridging was scored as 1.

When the intense regions were selected, the regions closest to the choriobasal unit in the marginal regions were preferred for counting. For subchorial counting the closest regions to chorial plaque, for midparenchymal counting the 1/3 sectional part of the placental thickness, and for basal counting the closest regions to the basal plaque were preferred. SPSS version 16.0 Statistical Analysis Software was used to analyze the relation between variants. Variations among groups were determined using the Mann-Whitney U test. The Kruskal-Wallis test was used for comparison of more than two groups. Probabilities of less than or equal to 0.05 were accepted as significant.

RESULTS

The clinical and pathological characteristics (the type of birth, the direction of cord turn, additional microscopic characteristics and villous maturation scores) of the 28 cases included in the study are demonstrated in Table I.

Villus counting from the previously defined regions, their number of included syncytial knots present, and the percentages of these are demonstrated in Table II. The group statistics are listed in Table III and Table IV. Mean parameters were calculated for every region. No significant difference between the mean parameters of the regions were demonstrated with Kruskal-Wallis test (p:0.148). No significant difference between the mean parameters of regions close to and away from cord entrance were found (p:0.101, and p:0.282, respectively). No significant differences between the mean parameters of marginal, subchorial, basal, and midparenchymal counting of regions close to and away from cord entrance were found (p:0,594, p:0,706, p:0,577, and p:0,577 respectively).

	Delivery route	Cord turn	Additional microscopical features	VMS		
1	VD	L	Х	22		
2	C/S	L	Х	23		
3	C/S	L	Focal stem villus calcification	22		
4	VD	L	Х	23		
5	VD	R	Cylindric change in amnion epithelium	22		
6	C/S	L	Subcorial fibrinoid excess	23		
7	C/S	L	Х	22		
8	C/S	R	Amniotic epithelium proliferation	22		
9	C/S	R	Х	22		
10	C/S	L	Amniotic mesencymal pigmented macrophages	23		
11	C/S	L	Х	22		
12	VD	L	Х	22		
13	C/S	L	Ductus vitellus remnant	22		
14	C/S	L	Х	22		
15	C/S	L	X			
16	C/S	L	Х	22		
17	VD	L	Х	22		
18	C/S	L	Intravillous fibrinoid excess	22		
19	C/S	L	Х	22		
20	C/S	L	Х	22		
21	C/S	L	Subcorial focal chronic infarct	22		
22	C/S	L	Х	22		
23	C/S	L	Х	22		
24	C/S	R	Х	22		
25	VD	R	Х	22		
26	C/S	R	Х	22		
27	C/S	R	X	22		
28	C/S	L	x	23		

Table I: The clinical and pathological characteristic of cases used in this study

C/S: Cesarean section, VD: Vaginal Delivery, VMS: Villous maturation score.

DISCUSSION

A syncytial knot, primarily defined under the light microscope, is defined as significant clustering of syncytial nuclei (Figure 5). Increase in syncytial knots and clustering of villus are defined as Tenney Parker Changes (TPC). TPC are primarily described as the determinant of preeclampsia. They are attributed to demonstrate placental ischemia later on. They have been associated with Factor 5 mutations lately (11,12). Syncytial knots are classified into three categories according to their mechanisms of formation. However, differentiation of these under the light microscope is quite difficult (1,13). Instead of being trophoblastic proliferations, most of the knots appear as slice artifacts of deformed villi due to villous angiogenesis, specifically during the last trimester. However, whatever their etiologies are, syncytial knots are known to demonstrate changes in trophoblastic turnover (3). Syncytial knots and bridging have various definitions in different studies. Baergen et al. have defined syncytial knots as clusters of 5 or more nuclei and only in terminal villi (14). However, it is reported that some of the counted villi do not meet the criteria and possess less than 5 nuclei. Syncytial bridging is described as one of the 4 etiological types of syncytial knots and the real bridging is described as a rare condition originating from fusion of 2 villi. The contact of the surfaces of the chorionic villi is designated as "touching" in the same study. Syncytial knots are defined as an increase in syncytiotrophoblastic nuclei, and the bridging as villous agglutination in the study of Redline et al. According to this, villous agglutination is

	2Bas	(%)	21.62	18.8	26.9	20	26.84	39.8	31.25	4.36	13.02	11.05	11.02	12.43	11.6	12.06	19.31	19.52	11.14	23.14	15.06	28.07	19	16.54	17.4	25.73	21.2	30.43	41.6	28.85	
	JDag	2Das	40/185	44/234	46/171	34/170	51/190	80/201	82/258	10/229	28/213	24/217	30/272	23/185	29/250	21/174	28/145	66/338	33/296	81/350	44/292	80/285	57/300	47/284	48/276	70/272	53/250	84/276	119/286	88/305	
	2Mid	(%)	13	25	16.49	22.6	17.03	25.58	13.49	8.62	15.05	16.24	8.06	22.74	7.05	5.55	14.44	22.18	12.06	18.72	42.81	23.8	23.5	20.48	22.07	19.33	15.83	43.75	30.72	29.72	
	PLAC	2 [V 110	26/200	65/260	47/284	66/292	31/182	55/215	34/252	22/255	28/186	32/197	25/310	58/255	18/255	13/234	40/277	65/292	38/315	53/283	149/348	70/294	110/468	68/332	51/231	99/512	38/240	70/160	106/345	132/444	
	2Sub	(%)	14.4	13.33	11.22	23.57	20	40.92	16.85	10.36	5.55	9.72	15	3.79	2.11	3.15	14.73	26.44	21.38	14.84	29.34	14.15	39.39	23.83	14.32	10	14.06	31.03	22.22	25.77	
	J5	ancz	18/125	16/120	32/285	33/140	40/200	97/237	30/178	20/193	9/162	18/185	24/160	11/290	5/237	6/190	28/190	64/242	50/197	53/357	54/184	63/445	117/297	87/360	48/335	30/300	37/263	63/203	110/495	150/582	- - -
	2X	(%)	12.19	12.13	17.6	26.38	17.6	24.69	9.82	11.36	29.7	11.89	15.95	19.84	26.37	12.93	6.84	17.85	20.74	30.56	58.33	30.98	16.12	19.3	38.42	25	28.02	38.66	34.7	29.28	
	λ¢	V7	25/205	33/275	44/250	95/360	25/142	41/166	17/173	25/220	60/202	27/227	30/186	51/257	67/254	30/232	10/146	50/280	56/270	118/386	175/300	110/355	60/372	50/259	78/203	87/348	44/157	116/300	76/219	82/280	
,	1Bas	(%)	12.94	13.23	13.93	18.75	14	15.21	17.91	13.79	5.84	17.78	9.46	7.03	13.77	8.45	5.71	29.26	20.21	25.37	29.14	18.45	22.11	36.2	25.21	25.11	30.07	29.04	30	20.38	
,	1 D 22	1 Das	18/139	36/272	23/165	51/272	28/200	28/184	24/134	40/290	18/308	37/208	16/169	14/199	42/305	12/142	18/315	96/328	76/376	85/335	109/374	62/336	69/312	105/290	30/119	58/223	77/256	97/334	75/250	84/412	
	1Mid	(%)	13.47	14.37	16.5	38.42	13.51	21.73	14.45	9.7	6.8	8.5	5.33	7.75	22.29	5	6.14	28.85	16.42	29.6	15.14	22.58	19.51	22.66	22.07	26.26	23.82	35.02	18.5	47.04	
	1 1/1:4	DIMIT	31/230	46/320	33/200	78/200	25/185	30/138	36/249	23/237	16/235	17/200	8/150	18/232	66/290	9/180	22/358	86/298	69/420	90/304	531/350	70/310	96/492	46/203	49/222	88/335	66/277	124/354	62/335	159/338	
	1Sub	(%)	8.17	5.76	12.27	9.38	19.67	20.58	8.16	7.14	15.33	7.92	20.94	10.3	10	5.43	8.49	42.72	10.4	17.25	29.18	13.85	30.34	25.84	39.72	20	17.93	20.88	18.55	23.34	-
	106	anet	13/159	12/208	27/220	38/405	24/122	42/204	20/245	11/154	25/273	13/165	40/191	20/194	30/300	10/184	18/212	91/213	26/250	64/371	61/209	29/214	88/291	69/267	58/146	131/655	47/262	47/225	77/415	102/437	•
	1X	(%)	15.56	3.54	15.43	35.58	17.85	28.7	21.25	17.72	25.27	32.86	13.49	9.13	36.95	4.61	9.61	37.58	29.18	47.56	36.39	37.72	15.29	18.06	51.13	33.43	23.65	18.98	21.17	27.27	
	1	VI	54/347	5/141	25/162	58/163	45/252	23/216	34/160	42/237	70/277	70/213	22/163	21/230	126/341	9/195	25/260	109/290	108/370	195/410	99/272	92/290	63/412	56/310	90/176	111/332	44/186	45/237	65/307	90/330	-
			1	2	3	4	5	9	7	8	6	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	;

Table II: The number of syncytiotrophoblastic knots and villi, their percentages (pct) counted at various areas

Area		n	Mean percentage	SD	Std. error mean	р	
x	1	28	24.46	12.24	2.31	0.504	
	2	28	22.97	11.17	2.11	0.394	
sub	1	28	17.12	9.78	1.84	0.70(
	2	28	17.55	9.96	1.88	0.706	
bas	1	28 18.86		8.31	1.57	0.577	
	2	28	20.63	8.89	1.68	0.5//	
mid	1 28		18.97	10.38	1.96	0.000	
	2	28	19.85	9.25	1.74	0.000	

Table III: Group Statistics; mean percentages of various areas and their comparison between peri-cord insertional and far from cord insertional zones

Table IV: Group Statistics; comparison between mean percentages of various areas (x, sub,bas, mid) within peri-cord insertional (1) and far from cord insertional (2) zones

Area		n	Mean percentage	SD	Std. error	р
	х	28	24.46	12.24	2.31	
	sub	28	17.12	9.78	1.84	
1	bas	28	18.86	8.31	1.57	0.101
	mid	28	18.97	10.38	1.96	
	Total	112	19.85	10.50	0.99	
	Х	28	22.97	11.17	2.11	
	Sub	28	17.55	9.96	1.88	
2	Bas	28	20.63	8.89	1.68	0.282
	Mid	28	19.85	9.25	1.74	
	Total	112	20.25	9.91	0.93	

described as clustering of the adjacent distal villi (>2, <20) inside fibrin and/or bridging (micro infarcts) at the syncytial knots (2). Others have mentioned that the formation of syncytial knots and bridging are slicing artifacts due to deformed villi with vascular proliferations, and the syncytial knots and the bridging are not defined as separate forms (1). We have scored one, two, or more villi including significant knot accumulation as 1 in our study. Syncytial knots or bridging are not differentiated. Some of these were proposed to be the contact regions described by Baergen et al. (14). The real and the pseudo bridging, and the differences from the contact regions should be clearly defined in future studies.

The determination of the region where the counting will take place is another argument. While in Handbook of Placental Pathology eds. Faye-Peterson OM, Heler DS, Joshi recommends counting from the midparenchymal region and from the knots on terminal villi (3), another study suggests the slice to be taken from the 75% of the basal part and from the knots on proximal stem villi and distal villi (2).



Figure 5: Syncytial Knots (Arrows) (H&E; x200).

Relative ischemia of marginal parts and the non-marginal parts of the basal and subchorial areas of the normal placentas have been reported (3). This thesis constitutes the basis of our study and our goals were to differentiate various areas according to their number of syncytial knots, and if there is a difference, to support the idea of relative ischemia and to contribute to the standardization of normal number of syncytial knots in various regions.

The theory of relative ischemia stems from physiological data about maternal circulation. The blood from the transformed arterioles entering the maternal circulation spouts into the subchorial region. This results in a fibrinoid accumulation defined as "subchorionic fibrinoid" and returns back to the basal area. It is reabsorbed from the open-ended veins (3). That is why the subchorial and the basal areas are the locations of blood accumulation. Another study has hypothesized that the collateral arterial circulation and the gestational arterial changes are fewer in the marginal regions (15). Besides, lesions like infarcts and x cell cysts that are classically known to be associated with hypoxia are reported to be more frequently seen in subchorial, basal and marginal regions.

No significant differences between the number of syncytial knots in any region were observed in our study. Although there is no significant increase in the number of syncytial knots in the marginal area or decrease in nonmarginal basal zones, these results should not be interpreted as nonexistence of relative ischemia, as the minor changes in perfusion may not effect the morphology. The increase in syncytial knot number is not the only finding of hypoxia and does not necessarily coexist with the other morphological changes of hypoxia. The morphological findings of hypoxia vary according to the degree and onset of hypoxia (16-19). For example, TPC may not accompany chorioangiosis that is thought to be formed due to long-lasting low grade hypoxia (3). Although we have excluded the cases with known hypoxia-associated changes from our study, there may be other findings yet not defined and associated with hypoxia. Future studies are needed to clarify this issue. We have an ongoing study proposed to demonstrate the number of syncytial knots from the same regions used in this study and investigate from where the knot increase has started, in cases with a villous maturation score of 32.

As a conclusion no differences between the number of syncytial knots counted from various regions of the normal term human placenta are demonstrated. The thesis of increased ischemia in marginal, non-marginal basal and non-marginal subchorial regions of the term, clinically and morphologically normal placentas are not supported in this study.

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