

Diagnostic Discrepancies Between Intraoperative Frozen Section and Permanent Histopathological Diagnosis of Brain Tumors

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ABSTRACT

Objective: Intraoperative frozen section (IOFS) diagnosis of brain tumors plays an important role in assessing the adequacy of the sample and determining the treatment plan. The aim of this study was to investigate the diagnostic accuracy between IOFS and permanent sections.

Material and Method: The authors reviewed the histopathological results of 383 brain tumors, including IOFS and permanent histological diagnosis. The cases were classified into three diagnostic compatibilities (i) Perfect fit; the diagnosis of IOFS was identical to the permanent diagnosis, (ii) Partial compatibility; IOFS diagnosis was not incorrect but was too broad to be considered full compatibility, (iii) Conflict; IOFS diagnosis is completely different from the permanent diagnosis. The permanent diagnosis was used as a primary criterion and was compared to IOFS diagnosis and recurrence rate using different statistical methods.

Results: 84% of the patients underwent craniotomy and tumor resection, while 15% only underwent tumor biopsy. Approximately, 53.8 % of the cases revealed perfect matching in the diagnosis between IOFSs and permanent sections, while 16.2% of the cases revealed complete mismatching in the diagnosis between the sections. The remaining 30% of the cases showed partial compatibility in the diagnosis between the two diagnostic methods. There was no significant difference in recurrence rate among all cases of different diagnostic compatibility ($p=0.54$).

Conclusion: There is a diagnostic discrepancy between IOFSs and permanent sections. However, cases that revealed no consensus in the diagnoses showed no negative effect on the patient outcome. Further studies should be conducted to explore the reasons of this conflict in the two diagnostic methods.

Keywords: Brain tumor, Histopathology, Frozen section, Diagnostic compatibility

INTRODUCTION

Brain tumors, known as intracranial tumors, are abnormal masses of tissue with cells that continuously grow and multiply. More than 150 types of brain tumors have been documented by the World Health Organization (WHO) (1). They can be categorized as primary and metastatic tumors. Primary tumors arise from brain tissue or surrounding structures, which include neuroglial cells, meninges, or the ependymal layer. They can be benign (low-grade) or malignant (high-grade). Secondary metastatic brain tumors include any organ cancer that hematogenously spreads to the brain. To distinguish primary from secondary brain tumors, a detailed clinical history and body imaging are

important to identify the origin of the tumor but not the exact histological subtype. Furthermore, intraoperative examination of tumor tissue is the gold diagnostic tool to differentiate the two types and to determine the surgical treatment plan.

Intraoperative frozen section (IOFS) diagnosis of brain tumors plays an important role in assessing the adequacy of the specimen, determining the surgical treatment plan, improving surgical procedures, and facilitating postoperative follow-up. In certain cases, when unexpected lesions cannot be identified by radiological imaging, the surgeon can determine the best procedure and endpoint of the operation (2,3). This can reduce the incidence of

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surgical complications and avoid unnecessary second surgical procedures. The key criteria and indications for requesting an intraoperative diagnosis by the surgeon are as follows: (a) intraoperative surgery would be affected by the diagnosis, (b) an unexpected lesion appears during surgery that is different from what was clinically suspected, (c) the primary goal is to obtain a biopsy diagnosis, and (d) the necessity to assess the margins if a total resection is planned (4-6). The distinction between primary tumor, lymphoma, metastatic tumor, or unusual lesions are considered the main reasons for requesting intraoperative diagnosis of brain tumors. Sometimes, the tumor is not accessible by surgery, and thus a stereotactic biopsy with IOFS is recommended. The assessment of brain tumors through IOFS is commonly used in clinical practice to verify the origin and type of tumor; however, the final diagnosis should be reported later after the permanent section (4,6).

Several studies have discussed the diagnostic accuracy of IOFS in assessing brain tumor type and grading, and inconsistencies were found between IOFS diagnosis and permanent diagnosis. Nevertheless, permanent paraffin-embedded examination remains the gold standard in diagnosing brain tumors. In Saudi Arabia, no publications have discussed the diagnostic accuracy between IOFS and permanent tissue diagnosis of brain tumors. In this study, we aimed to assess the compatibility between the results of frozen sections and permanent sections in patients diagnosed with brain tumors. We also discussed the reasons attributed to the lack of diagnostic accuracy between the two diagnostic methods.

MATERIALS and METHODS

Case Stratification

In this retrospective study, we reviewed the histopathological reports of 383 primary and secondary brain tumors between 2013 and 2019 from two medical institutions in Saudi Arabia. The study was ethically approved by the National Biomedical Ethics Committee at King Abdulaziz University (HA-02-J-008) under general ethical approval. Patient age, gender, intraoperative and permanent histological diagnosis and grading, and recurrence rate were used as statistical factors. The permanent section result was used as a primary criterion, and the frozen section diagnosis was compared with the final diagnosis.

Histopathological Confirmation

The histopathological report of IOFS and permanent paraffin-embedded section of each patient with a brain tumor was examined. The diagnostic compatibility between

the two diagnostic methods was determined on the basis of tumor type and grading. The cases were classified into three degrees of diagnostic compatibility as follows: 1) the diagnosis of IOFS was identical to the final diagnosis (perfect fit); 2) the diagnosis of IOFS was not incorrect but was too broad to be considered fully compatible (partial compatibility); and 3) the diagnosis of IOFS were incorrect and differs from the final diagnosis (conflict).

Statistical Analysis

Statistical analysis was performed using SPSS software (version 20). To describe the data, we used means, frequencies, and percentages. To assess the diagnostic compatibility of different methods, we used the linear model, ANOVA and people's Chi-square test. The number and types of discrepancies were identified.

RESULTS

The mean age of the 383 patients was 37.6 ± 23.1 years (females: 212 (55.4%); males: 171 (44.6%)). 84.86% of the patients underwent craniotomy and tumor resection, while 15.14% only underwent tumor biopsy (open or stereotactic biopsy). Most of the tumors were grade IV (n=177, 46.2%), followed by grade II (n=105, 27.4%) (Table I). Tumour locations were summarized in (Figure 1).

Table I: Descriptive distribution of the data in this study.

	Overall (n=383)
Age	
Mean (SD)	37.6 (23.1)
Range	1.0-87.0
Gender	
Female	171 (44.6%)
Male	212 (55.4%)
Type of Tissue Removal	
Tumor biopsy	58 (15.14%)
Surgical resection	325 (84.86%)
Diagnostic Compatibility	
Conflict	62 (16.2%)
Partial Compatibility	115 (30.0%)
Perfect fit	206 (53.8%)
WHO Grading	
Grade I	105 (27.4%)
Grade II	53 (13.8%)
Grade III	36 (9.4%)
Grade IV	177 (46.2%)
Unclassified	12 (3.1%)

Diagnostic Compatibility Between IOFS and Permanent Sections

The diagnostic compatibility between IOFS and permanent section showed comprehensive variability, as 53.8% (n=206) of the cases showed perfect match between IOFS and permanent section diagnosis, while 16.2% (n=62) of the cases showed complete mismatch. Furthermore, 30% (n=115) of the cases showed partial compatibility between the two diagnostic methods (Table I; Figure 2). For example, the cases diagnosed as glioblastoma with IOFS were also diagnosed as glioblastoma with permanent section; however, these cases represented 13% perfect compatibility among all investigated brain tumors (Table II). In a few cases, glioblastomas were misdiagnosed with IOFS as low-grade gliomas, necrotic cells, or glioneuronal tumors (Table III).

Diagnostic Compatibility of Low-Grade and High-Grade Gliomas

Approximately 9% of the cases diagnosed as low-grade gliomas with IOFS were diagnosed as high-grade gliomas with permanent sections, and one case was found to be a meningioma (Table II). Conversely, 17% of the cases diagnosed as high-grade gliomas with IOFS were diagnosed on permanent sections as either low-grade gliomas or a

different histological subtype such as medulloblastoma or metastasis. Furthermore, 19% of the cases diagnosed with infiltrating glioma with IOFS showed partial compatibility on permanent sections. The final diagnosis was grade II, such as cases of oligodendrogliomas (Table II).

Diagnostic Compatibility Between Atypical, Reactive, and Necrotic Cells

From the 22 cases diagnosed as atypical glial cells with IOFS, two of them were permanently diagnosed as mature teratoma and hemangioblastoma. The remaining 20 cases of atypical glial cells were compatible with the same glioma histogenesis, regardless of the grading. These cases were considered partially compatible. The cases that were showing reactive cells in IOFS (n=6, 9.6%) were diagnosed differently from permanent sections (Table II). These cases included teratoma, hemangioblastoma, germinoma, ganglioglioma, pilocytic astrocytoma, and pituitary adenoma. Cases for which IOFS revealed necrotic cells were diagnostically deferred at the time of surgery. Their permanent sections revealed malignant tumors except one case (pilocytic astrocytoma in the brainstem), which was re-diagnosed as diffuse midline glioma.

Relationship Between Recurrence Status and Diagnostic Compatibility

There was no statistically significant relationship between the recurrence rates and the diagnostic compatibilities of different brain tumors (p=0.54) (Table IV). Cases that showed conflicts in diagnostic compatibility showed no significant differences in recurrence rates compared to cases with perfect diagnosis matches. This clarifies that conflict in the diagnostic compatibility between IOFS, and permanent histological sections may not affect tumor recurrence rate.

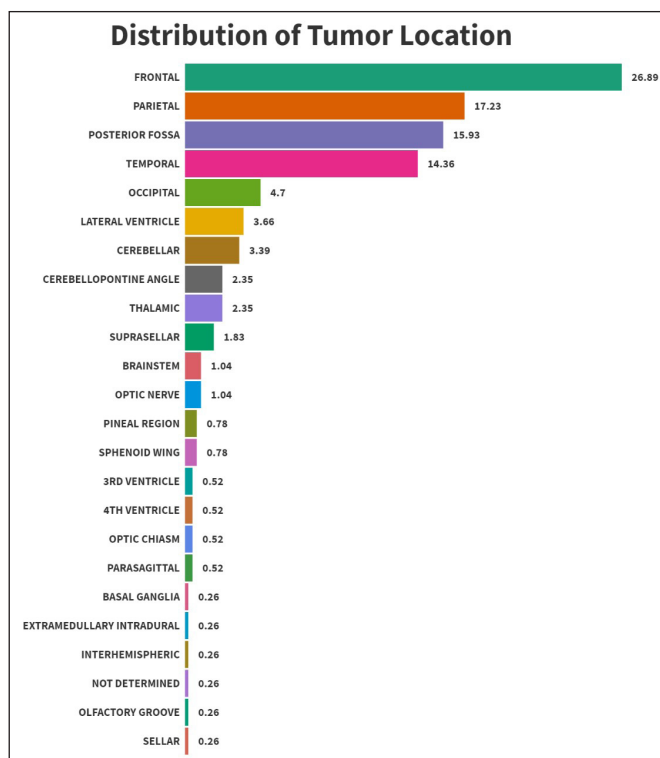


Figure 1: Tumor locations of all cases enrolled in this study. There is 0.25% of cases did not have specific location or the location was not mentioned in the report.

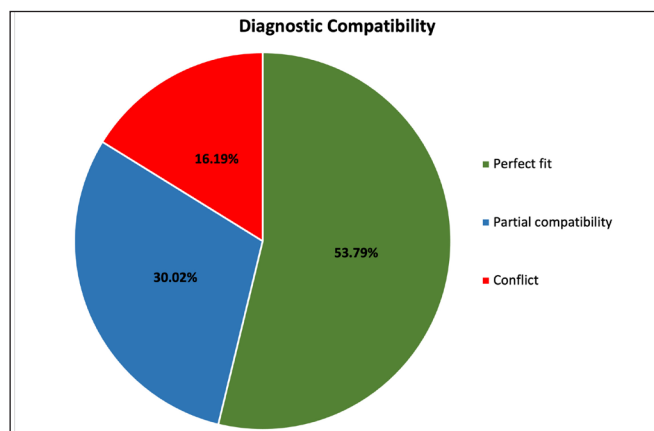


Figure 2: Diagnostic compatibility between intraoperative frozen sections and permanent paraffin-embedded sections in all brain tumors enrolled in this study.

Table II: Diagnostic compatibilities of the probabilities of all frozen sections compared with permanent final diagnosis in brain lesions. This table compares the diagnostic compatibility of each tumor through the total number of all cases.

Frozen Section Diagnosis, n(%)	Conflict (n=62)	Partial compatibility (n=115)	Perfect fit (n=206)	Total (n=383)
Atypical glial cells	2 (3.2)	20 (17.4)	0 (0.0)	22 (5.7)
Ependymoma	1 (1.6)	0 (0.0)	8 (3.9)	9 (2.3)
Glioblastoma	0 (0.0)	0 (0.0)	28 (13)	28 (7.3)
Glioma	0 (0.0)	1 (0.9)	0 (0.0)	1 (0.3)
Glioneuronal tumor	3 (4.8)	3 (2.6)	1 (0.5)	7 (1.8)
Hemangioblastoma	0 (0.0)	0 (0.0)	5 (2.4)	5 (1.3)
Hemorrhage	2 (3.2)	0 (0.0)	0 (0.0)	2 (0.5)
High grade glioma	17 (27.4)	3 (2.6)	101 (49)	121 (31)
Infiltrating glioma	2 (3.2)	19 (16)	2 (1.0)	23 (6.0)
Low grade glioma	6 (9.7)	61 (53)	10 (4.9)	77 (20)
Lymphoma	2 (3.2)	0 (0.0)	0 (0.0)	2 (0.5)
Malignant cells	0 (0.0)	3 (2.6)	0 (0.0)	3 (0.8)
Meningioma	1 (1.6)	2 (1.7)	27 (13)	30 (7.8)
Mesenchymal tumor	0 (0.0)	2 (1.7)	0 (0.0)	2 (0.5)
Metastatic tumor	0 (0.0)	0 (0.0)	9 (4.4)	9 (2.3)
Necrotic cells	20 (32.3)	1 (0.9)	0 (0.0)	21 (5.5)
Pilocytic astrocytoma	0 (0.0)	0 (0.0)	10 (4.9)	10 (2.6)
Reactive cells	6 (9.7)	0 (0.0)	0 (0.0)	6 (1.6)
Schwannoma	0 (0.0)	0 (0.0)	5 (2.4)	5 (1.3)

Table III: List of brain tumors with their misdiagnosis with intraoperative frozen sections (IOFS).

Final diagnosis	Misdiagnosis in IOFS
Glioblastoma	Low-grade glioma (n=4) Glioneuronal tumor (n=2) Necrotic cells (n=14)
Metastatic carcinoma	High-grade glioma (n=1)
Medulloblastoma	High-grade glioma (n=5) Necrotic cells (n=4)
Diffuse large B-cell lymphoma	High-grade glioma (n=1) Infiltrating glioma (n=1)
Pilocytic astrocytoma	High-grade glioma (n=1) Necrotic cells (n=1) Lymphoma (n=1) Reactive cells (n=1)
Schwannoma	Meningioma (n=1)
Ependymoma	High-grade Glioma (n=1)

DISCUSSION

Various types of brain tumors have been documented by the 2016 WHO-Central Nervous System (1). They are categorized as primary and metastatic tumors. Primary tumors arise from brain tissue or surrounding structures and are benign (low-grade) or malignant (high-grade). Secondary metastatic brain tumors arise from any systemic organ cancer. To distinguish primary from secondary brain tumors, body imaging and intraoperative examination of tumor tissue are considered the best diagnostic methods, which can also determine the surgical treatment plan. Our current study included 369 primary brain tumors and 14 metastatic tumors to the brain.

The IOFS analysis was first introduced in 1891 by a pathologist at Johns Hopkins Hospital, and is currently used worldwide. The application of this method became much easier through improvements in cryostat devices (7). IOFS is mainly useful for solid and stretchy tumors such as meningiomas, schwannoma, and most metastases in which cell smears are difficult to prepare (8,9). It also provides

Table IV: Relationship between recurrence rate and diagnostic compatibilities of intraoperative frozen sections and permanent sections.

	No recurrence (n=216)	Recurrence (n=167)	Total (n=383)	p value
Diagnostic Compatibility, n(%)				0.547 ¹
Conflict	36 (16.7)	26 (15.6)	62 (16.2)	
Partial Compatibility	69 (31.9)	46 (27.5)	115 (30.0)	
Perfect fit	111 (51.4)	95 (56.9)	206 (53.8)	

¹ Linear model ANOVA

good architectural detail of the lesion, and better reveals histological patterns and cell morphology (10,11). IOFS is commonly used to verify the competence, and origin of brain tumors (4,6).

There are two methods to assess the diagnostic compatibility between IOFS and permanent sections. The first method is to determine whether the IOFS diagnosis matches the permanent diagnosis, e.g., glioblastoma diagnosed with IOFS is also diagnosed as glioblastoma with permanent section. The second method is to determine whether the permanent diagnosis is histologically close to the IOFS diagnosis, e.g., glioblastoma diagnosed on permanent section is observed as atypical glial cells in frozen sections. In our current study, both methods were used; however, we focused more on the first method as it represents the foundation of the general diagnostic accuracy of brain tumors.

The compatibility between IOFS and permanent section in clinical practice always shows diagnostic diversity. In our study, 53% of the lesions diagnosed on permanent histological sections matched the IOFS diagnosis, while 16% showed complete mismatching (Figure 2). This variation is considered clinically significant as it reveals that IOFS itself should not be solely used as a definitive diagnostic method. Most of the discordances between IOFS and permanent histological sections were observed in high-grade gliomas. This might be attributed to the difficulty in the microscopic interpretation, and the lack of experience. It is sometimes quite hard to identify the cytoplasmic processes in the frozen section, which makes it difficult to differentiate glial background from other types of tumors, particularly metastasis. For example, glioblastomas may be misdiagnosed as low-grade gliomas during the IOFS. This is because the peripheral edges of glioblastoma may contain reactive or atypical glial cells without necrosis or endothelial proliferation. Furthermore, the distinction between high-grade gliomas and metastasis is also difficult unless the malignant glial cells with gliotic background are clearly seen. In diffusely astrocytic cases, missing IOFS diagnosis of astrocytomas could be due to the thickness

of the sections and technical problems with staining, which disrupts the cellular morphology. The difficulties in diagnosing hemangioblastoma with IOFS were because most hemangioblastomas are bloody and vascular, which may mask the cellular features of the tumors. Cases that showed reactive or necrotic cells with IOFS revealed different diagnostic results on permanent sections. For example, most of the cases diagnosed as necrotic cells with IOFS had a permanent section diagnosis of glioblastoma, while IOFS diagnosis of reactive cells had a permanent section diagnosis of teratoma, germinoma, ganglioglioma, pilocytic astrocytoma, or pituitary adenoma. These findings depend on the biopsy location, and the size of the necrotic component in the tumor.

The diagnostic accuracy in our study was 53%. In other studies, the diagnostic accuracy was 92.4% in Talan-Hraniloviæ et al. (12), and 95% in Roessler et al. (13) with 89% complete concordance among 4,172 patients. The most accurate IOFS diagnoses in Roessler's study were made in cases of meningioma (97.9%), metastasis (96.3%), and glioblastoma (95.7%) (13). Although the high-grade gliomas showed high discordance between IOFS and permanent histological section, the diagnostic compatibility was still acceptable. Plesec et al., had almost 3% discordancy among 2,156 cases. Approximately 80% of these cases were spindle cell lesions, astrocytoma versus oligodendroglioma, lymphoma, reactive versus neoplastic process, and tumor overgrading (14). We also have found that 97% of meningioma cases showed perfect matching between the IOFS and permanent section diagnoses. One case turned out to be a schwannoma.

Several reasons might be attributed to this mismatching. Theoretically, the reasons can either be related to the quality of surgical procedure or the quality beyond the laboratory diagnostic techniques. If the neurosurgeon takes the biopsy from the necrotic part of the tumor or from the surrounding reactive area, the tissue may be fragmented, nonviable, or nondiagnostic. An experienced neurosurgeon should navigate the tumor using a brain imaging navigator before taking the biopsy. However, multiple punch biopsies

are recommended in cases where the necrotic parts are deeply seated in the tumour center. Freezing processes may also mask the tissue diagnosis. Calcification, autolysis, improper hematoxylin and eosin staining, or inappropriate usage of hemostatic agents all can contribute to incorrect diagnosis.

As we mentioned before, one of the common reasons for diagnostic disagreements between IOFS, and permanent paraffin-embedded section is the microscopic interpretation. Most pathologists who see the brain tumors during IOFS are non-neuropathologists with limited experience in the brain. Pathologists, who see brain tumors during IOFS, must have enough knowledge to correlate between brain imaging and histology. This would help pathologists minimize the differential diagnoses, and approximate the diagnostic probabilities with neurosurgeons. This problem is more common in developing countries where health care systems have shortage of neuropathologists. A fully certified neuropathologist with a minimum two-year training in neuropathology will have better diagnostic outcome than a non-neuropathologist.

CONCLUSION

The role of frozen sections during intraoperative consultation is important. Our results showed some diagnostic discrepancies between the intraoperative diagnosis of brain tumors, and permanent final diagnosis. Appropriate knowledge of pathologists regarding radiological findings and microscopic interpretation with proper communication with neurosurgeons are required to minimize IOFS misdiagnosis. Further studies should be conducted to determine the reasons for this discrepancy and to solve the problems related to this incompatibility.

CONFLICT of INTEREST

The authors declare no conflicts of interest.

AUTHORSHIP CONTRIBUTIONS

Concept: **MK**, Design: **MK**, Data collection or processing: **YM, AB, RS, AL, TS, AS**, Analysis or Interpretation: **MK, SB, SH**, Literature search: **MK, SB, RS**, Writing: **MK, SB, YM, RS, AS**, Approval: **MK, SB, AL**.

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