The importance of β -catenin during melanoma formation

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 β -Catenin has an important role in the differentiation of melanocytes and progression to melanomas. This protein is involved in cell-cell adhesion, cell signaling and gene transcription that are disrupted during malignant transformation. β -Catenin has a cell-cell adhesion function mediated by cadherins. The intercellular signaling is regulated with glycogen synthase 3β (GSK 3β)/Wnt pathway and the gene transcription activity is mediated by DNA-binding LEF/TCF proteins. The oncogenic activity of β -catenin is directly associated with its participation in multiple protein-protein interactions and its role during gene expression. The microphthalmia-associated transcription factor (MITF), a target for β -catenin, has a role in melanoblast proliferation and in its differentiation. In this review β -catenin interactions and targets that are involved in the differentiation of melanocytes and its particular role in the expression of MITF are described.

Keywords: β-catenin, cadherin, microphthalmia-associated transcription factor, cell-cell adhesion, melanocyte

Development of melanocyte and interaction with keratinocyte

Melanoblasts are derived from neural crest cells. These cells migrate along the dorso-lateral pathway, between the somites and the ectoderm, then they cross the basement membrane separating the dermis from the epidermis, and invade the epidermis, afterwards they are called melanocytes once they produce melanin.¹ The melanocyte in human skin is normally embedded in the basal layer of keratinocytes. Interactions between melanocytes and keratinocytes are thought to be important in the control of melanocyte growth.² Human melanocytes and keratinocytes express E-cadherin, which mediates functional their heterotypic interactions.^{3–5} The cell-cell adhesion function of E-cadherin depends on the presence of cytoplasmic proteins that include α , β and γ catenin. Disruption of the expression of E-cadherin and

catenins has been implicated in tumor progression and metastasis.⁶ Melanocytes may transform to melanoma, in some cases after forming a nevus and progresses to radial growth phase and to vertical growth phase. Melanocytes, but not nevus cells or melanoma cells express E-cadherin. During the malignant transformation, the classical melanocyte-keratinocyte interaction is lost for a homotypic melanocyte-melanocyte interaction in which cell-cell adhesion molecules are involved and affected.^{6,7}

The main proteins involved in cell-cell adhesion are those of the cadherin/catenin complex that have a key role during normal and pathological differentiation of melanocytes. We shall discuss some interactions of these molecules with β -catenin in the progression of human melanoma.

Functions of β -catenin in different cellular compartments

Depending on its localization and its interactions, β catenin is involved in cell-cell adhesion, cell signaling and gene transcription. The first known function of β catenin is in cell-cell adhesion. Cadherins are anchored within cells by dynamic associations with catenins, which link them to the actin filaments. Cadherins constitute a superfamily of Ca²⁺-dependent cell-cell adhesion molecules. They are transmembrane proteins composed of an extracellular domain, a membranespanning region, and a highly conserved cytoplasmic part. The cytoplasmic domain of cadherins directly interact with either β -catenin / a-catenin and γ -catenin (=plakoglobin)/a-catenin bound to actin filaments via alfa-catenin.⁸ In normal epidermis, β-catenin expression and its cellular localization together with Ecadherins can be shown by their membranous immunohistochemical staining.

β-catenin if not incorporated in cell-cell adherens junctions is found as a large complex. It includes the protein encoded by the APC tumor suppressor gene, the GSK-3ß serine/threonine protein kinase and axin/conductin protein.9-12 GSK3ß can phosphorylate serine/threonine residues and consequent degradation of β -catenin by the ubiquitin-proteosome system. Activation of the Wnt/Wg pathway block this degradation which regulate the amount of free cytoplasmic β-catenin. Increased amount of free βcatenin translocate into the cell nucleus. Nuclear βcatenin may interact with members of the TCF/LEF family of transcription factors and stimulate transcription of a various target genes.^{13–15} Loss or gain of transcription factor gene function can notably play a major role in tumor growth and metastasis of melanoma.¹⁶ These genes are involved in cell proliferation such as c-Myc, cyclin D1 or MITF promoters.^{16–18} The importance of these different proteins in the biology of the melanocytes is not vet fully appreciated. A critical role played by the Wnt/ β catenin signaling pathway in the progression of melanoma is due to altered equilibrium of different complexes β-catenin in various cellular compartments.9-11

MITF expression and its role in melanoma progression

MITF, is a member of bHLH-Zip family transcription factor, it may either decrease or increase gene transcription. In humans, MITF mutation is characterized by hearing deficiencies, white forelock and ocular anomalies (Waardenburg syndrome, Type II).¹⁹ Human MITF is homologous to mouse micropthalmia gene (mi). Among the various MITF isoforms, the M isoform is found in melanocytes. Melanocyte cell differentiation and function are regulated by a wide spectrum of genes among which MITF has a key role. During embryogenesis, MITF is involved in survival and proliferation of melanoblast. In differentiated melanocytes, it regulates the expression of melanocyte differentiation markers, such as tyrosinase.¹⁶ Normal skin melanocyte, nevi, dysplastic nevi and metastatic tumors are detected with nuclear staining using an antibody against human MITF.²⁰ It appears that MITF expression is a molecular marker for diagnosis and its presence has a good prognosis in melanomas.²⁰⁻²² There is also a study suggesting that MITF may be a prognostic marker in patients with intermediate-thickness melanoma.²² The MITF gene regulation depends at least on Sox10, Pax3 and β -catenin/TCF.¹⁷

Oncogenic activation of β -catenin

During the transformation of melanocytes to melanomas, β-catenin is no longer mainly located at the cell surface, but rather accumulated in the cytoplasm and may signal directly to the nucleus.^{23–26} Aberrant β -catenin expression, with significant loss of membranous expression lead to metastasis of melanoma was also demonstrated previously.²⁷⁻²⁸ It is likely that stabilization and overexpression of β catenin, either as a result of mutation in β -catenin itself or other components of the Wnt signaling pathway play an important role in melanoma progression (Figure 1). Many of those colon cancer with wild-type APC are seen after mutation of the β -catenin gene.^{29,30} In a limited number of melanoma cases, point mutations at the level of the serine,^{37,45} and truncated form of B-catenin in its N-terminal region were identified.31,32 Note, however, that the presence of mutations in melanoma cell lines is higher than in

melanoma cases, although there is an accumulation of β-catenin at nucleus.^{28,31,32} One possibility, among others yet to be tested, is the increased frequency of mutations during cell culture. Another hypothesis is β catenin mutation, which is already present in some cutaneous melanoma cells that might facilitate their adaptation to the culture media.³³ A non-functional APC may also fail to export nuclear β-catenin to the cytoplasm.³⁴ The consequence of either APC inactivation or β-catenin mutation is similar: a failure to degrade β -catenin properly, also aberrant function of may result β-catenin in increased cellular proliferation.^{17,35} Upregulation of β -catenin induces the Lef/Tcf transcription factors and lead to continuous

Conclusions

 β -catenin, once activated with or without mutation may constitutively interact with one or more targets related to cell growth, cell proliferation, cell invasion and apoptosis.^{37,38} Melanoma cell lines indicate that regulation of melanoma growth by the Wnt β -catenin / Lef pathway is functionally dependent on the MITF transcription factor.^{18,39,40} It is possible, that MITF might also participate in β -catenin-mediated growth, invasion and pigmentation control in melanoma cases.²⁰

Although Wnt/β -catenin signaling is required early in melanocyte differentiation, possibly through



Normal melanocyte

Transformed melanocyte

Figure 1: Small pool of β -catenin is bound to the APC–axin-containing protein complex, which targets β -catenin for degradation. In malignant transfomation of melanocyte, active pools of β -catenin in the cytoplasm and nucleus interact with TCF-type transcription factors and stimulate the expression of genes that are targets of the Wnt signaling pathway.

activation of target genes. Altogether, it is not surprising that Myc and MITF, two targets of β -catenin, are frequently not properly regulated in melanomas.^{13,17,36}

the ability to activate MITF expression, it is not yet clear whether β -catenin is required at later times. β -Catenin-mediated activation of MITF expression is currently emphasized, however other genes may also play a primary role in mediating the response of melanocytes to Wnt/ β -catenin signaling.¹⁶ β -catenin and its target genes related with Wnt/ β -catenin signaling pathway have great importance at melanocyte lineage. The nature of those target genes that may contribute to malignant transformation must be investigated in future studies.

References

- Larue L, Kumasaka M, Goding CR. β-catenin in the melanocyte lineage. Pigment Cell Res 2003;15:1-6.
- 2. Hirobe T. Keratinocytes are involved in regulating the developmental changes in the proliferative activity of mouse epidermal melanoblasts in serum-free culture. Dev Biol 1994;161:59-69.
- Tang A, Eller MS, Hara M, Yaar M, Hirohashi S, Gilchrest BA. E-cadherin is the major mediator of human melanocyte adhesion to keratinocytes in vitro. J Cell Sci 1994;107:983-992.
- Cowley GP, Smith MEF. Cadherin expression in melanocytic naevi and malignant melanomas. J Pathol 1996;179:183-187.
- Jouneau A, Yu YQ, Pasdar M, Larue L. Plasticity of cadherincatenin expression in the melanocyte lineage. Pigment Cell Res 2000;13:260-272.
- Li G, Herlyn M. Dynamics of intercellular communication during melanoma development. Mol Med Today 2000;6:163-169.
- Herlyn M, Berking C, Li G, Satyamoorthy K. Lessons from melanocyte development for understanding the biological events in naevus and melanoma formation. Melanoma Res 2000;10:303-312.
- Silye R, Karayiannakis AJ, Syrigos KN, Poole S, van Noorden S, Batchelor W, Regele H, Sega W, Boesmueller H, Krausz T, Pignatelli M. E-cadherin/ catenin complex in benign and malignant melanocytic lesions. J Pathol 1998;186:350-355.
- 9. Moon RT, Bowerman B, Boutros M, Perrimon N. The promise and perils of Wnt signaling through beta-catenin. Science 2002;296:1644-1646.
- Gottardi CJ, Gumbiner BM. Adhesion signaling: how betacatenin interacts with its partners. Curr Biol 2001;11:R792-R794.
- Ben-Ze'ev A. The dual role of cytoskeletal anchor proteins in cell adhesion and signal transduction. Ann N Y Acad Sci 1999;886:37-47.
- Polakis P. The oncogenic activation of beta-catenin. Curr Opin in Genet Dev 1999;9:15-21.
- He TC, Sparks AB, Rago C, Hermeking H, Zawel L, da Costa LT, Morin PJ, Vogelstein B, Kinzler KW. Identification of c-MYC as a target of the APC pathway. Science 1998;281:1509-1512.
- Shtutman M, Zhurinsky J, Simcha I, Albanese C, D'Amico M, Pestell R, Ben-Ze'ev A. The cyclin D1 gene is a target of the beta-catenin/LEF-1 pathway. Proc Natl Acad Sci U S A 1999;96:5522-5527.
- Tetsu O, McCormick F. Beta-catenin regulates expression of cyclin D1 in colon carcinoma cells. Nature 1999;398:422-426.
- Nyormoi O, Bar-Eli M. Transcriptional regulation of metastasisrelated genes in human melanoma. Clin Exp Metastasis 2003;20:251-263.

- 17. Goding CR. MITF from neural crest to melanoma: signal transduction and transcription in the melanocyte lineage. Genes Dev 2000;14:1712-1728.
- Widlund HR, Horstmann MA, Price ER, Cui J, Lessnick SL, Wu M, He X, Fisher DE. Beta-catenin-induced melanoma growth requires the downstream target Microphthalmia-associated transcription factor. J Cell Biol 2002;158:1079-1087.
- Mouriaux F, Vincent S, Kherrouche Z, Maurage CA, Planque N, Monte D, Labalette P, Saule S. Microphthalmia transcription factor analysis in posterior uveal melanomas. Exp Eye Res 2003;76:653-661.
- King R, Weilbaecher KN, McGill G, Cooley E, Mihm M, Fisher DE. Microphthalmia transcription factor. A sensitive and specific melanocyte marker for melanoma diagnosis. Am J Pathol 1999;155:731-738.
- Dorvault CC, Weilbaecher KN, Yee H, Fisher DE, Chiriboga LA, Xu Y, Chhieng DC. Microphthalmia transcription factor: a sensitive and specific marker for malignant melanoma in cytologic specimens. Cancer 2001;93:337-343.
- 22. Salti G, Manougian T, Farolan M, Shilkaitis A, Majumdar D, Das Gupta TK. Microphthalmia transcription factor: a new prognostic marker in intermediate-thickness cutaneous malignant melanoma. Cancer Res 2001;60:5012-5016.
- Sanders DS, Blessing K, Hassan GA, Bruton R, Marsden JR, Jankowski J. Alterations in cadherin and catenin expression during the biological progression of melanocytic tumours. Mol Pathol 1999;52:151-157.
- Aberle H, Schwartz H, Kemler R. Cadherin-catenin complex: protein interactions and their implications for cadherin function. J Cell Biochem 1996;61:514-523.
- Hsu MY, Meier F, Herlyn M. Melanoma development and progression: a conspiracy between tumor and host. Differentiation 2002;70:522-536.
- Barker N, Clevers H. Catenins, wnt signaling and cancer. Bioessays 2000;22:961-965.
- Omholt K, Platz A, Ringborg U, Hansson J. Cytoplasmic and nuclear accumulation of beta-catenin is rarely caused by CTNNB1 exon 3 mutations in cutaneous malignant melanoma. Int J Cancer 2001;92:839-842.
- Rimm DL, Caca K, Hu G, Harrison FB, Fearon ER. Frequent nuclear/cytoplasmic localization of beta-catenin without exon 3 mutations in malignant melanoma. Am J Pathol 1999;154:325-329.
- Morin PJ, Sparks AB, Korinek V, Barker N, Clevers H, Vogelstein B, Kinzler KW. Activation of beta-catenin-Tcf signaling in colon cancer by mutations in beta-catenin or APC. Science 1997;275:1787-1790.
- 30. Ilyas M, Tomlinson IP, Rowan A, Pignatelli M, Bodmer WF. Beta-catenin mutations in cell lines established from human colorectal cancers. Proc Natl Acad Sci USA 1997;94:10330-10334.
- Rubinfeld B, Robbins P, El-Gamil M, Albert I, Porfiri E, Polakis P. Stabilization of beta-catenin by genetic defects in melanoma cell lines. Science 1997;275:1790-1792.
- 32. Robbins PF, El-Gamil M, Li YF, Kawakami Y, Loftus D, Appella E, Rosenberg SA. A mutated beta-catenin gene encodes a melanoma-specific antigen recognized by tumor infiltrating lymphocytes. J Exp Med 1996;183:1185-1192.
- 33. Demunter A, Libbrecht L, Degreef H, De Wolf-Peeters C, van den Oord JJ. Loss of membranous expression of beta-catenin is associated with tumor progression in cutaneous melanoma and rarely caused by exon 3 mutations. Mod Pathol 2002;15:454-461.

- Henderson BR, Galea M, Schuechner S, Leung L. Lymphoid enhancer factor-1 blocks adenomatous polyposis coli-mediated nuclear export and degradation of beta-catenin. Regulation by histone deacetylase 1. J Biol Chem 2002;277:24258-24264.
- 35. Peifer M. Beta-catenin as oncogene: the smoking gun. Science 1997;275:1752-1753.
- Takeda K, Yasumoto K, Takada S, Watanabe K, Udono T, Saito H, Takahashi K, Shibahara S. Induction of melanocyte-specific microphthalmia-accociated transcription factor by Wnt-3a. J Biol Chem 2000;275:14013-14016.
- Orsulic S, Huber O, Aberle H, Arnold S, Kemler R. E-cadherin binding prevents beta-catenin nuclear localization and betacatenin/LEF-1-mediated transactivation. J Cell Sci 1999;112:1237-1245.
- 38. Kageshita T, Hamby CV, Ishihara T, Matsumoto K, Saida T, Ono T. Loss of beta-catenin expression associated with disease progression in malignant melanoma. Br J Dermatol 2001;145:210-216.
- 39. Aberdam E, Bertolotto C, Sviderskaya EV, de Thillot V, Hemesath TJ, Fisher DE, Bennett DC, Ortonne J-P, Ballotti R. Involvement of Microphthalmia in the inhibition of melanocyte lineage differentiation and of melanocgenesis by agouti signal protein. J Biol Chem 1998;273:19560-19565.
- Yasumoto K, Takeda K, Saito H, Watanabe K, Takahashi K, Shibahara S. Microphthalmia-associated transcription factor interacts with LEF-1, a mediator of Wnt signaling. EMBO J 2002;21:2703-2714.